

British Journal of Pharmacy

www.bjpharm.hud.ac.uk

Proceedings of the 14th APS International PharmSci 2023

Intranasal Vaccine Formulation: Advancing Towards Nasal Dry Powder Formulation

K. Forkuoh¹, L. Urbano¹, D. Murnane¹, L. Kerai^{1*}

¹Centre for Topical Drug Delivery and Toxicology, School of Life and Medical Sciences, University of Hertfordshire, Hatfield AL10 9AB, UK

ARTICLE INFO	S U MM A R Y									
Received: 09/08/2023 Revised: 17/08/2023 Accepted: 18/08/2023 Published: 30/12/2023	Unlike conventional mRNA vaccines, intranasal vaccines display several advantages, including the ability to generate strong mucosal immunity. However, the formulation challenges associated with intranasal mRNA vaccines have so far hindered their extensive application and further research in the area of formulation									
*Corresponding author. E-mail:l.kerai-varsani @herts.ac.uk	development and optimisation are required before translation. This study aims to optimise the manufacturing conditions of inhalable dry powders by examining the impact of temperature and lipid composition on liposome particle size and the effect of spray-drying parameters on the particle size of spray-dried powders. The									
KEYWORDS: Liposomes, microfluidics, spray-drying, vaccines	results indicate that liposomes generated at 20°C exhibited notably larger sizes than those produced at 55°C, irrespective of the lipid composition. Notably, dry powder formulations featuring larger particle sizes were achieved at a low gas flow rate of 25mm.									
	<u> </u>									

BY 4.0 Open Access 2023 – University of Huddersfield Press

INTRODUCTION

Recently, Lipid nanoparticle-mRNA vaccines, Pfizer-BioNTech and Moderna COVID vaccines have demonstrated a remarkable ability to elicit potent immune systemic responses against SARS-CoV-2 ^{1,2}. However, their inability to elicit a robust mucosal immunity against lung diseases is a disadvantage ³.

Intranasal administration of lipid nanoparticle vaccines has emerged as a promising and noninvasive option for mucosal delivery for generating a strong mucosal immunity against respiratory pathogens 4. Stabilizing mRNA with dry powderbased mucoadhesive formulations can ensure dosing extended stability, improved accuracy, nasal residence time and patients' acceptability 5. However, the formulation challenges associated with intranasal mRNA vaccines have so far hindered their extensive application and further research in the area of formulation development and optimisation are required before translation 6.

This study aims to optimise the manufacturing conditions of inhalable dry powders by examining: 1) The impact of temperature and lipid composition on liposome particle size 2) The effect of spray-drying parameters (e.g. gas flow, inlet temperature and feed rate) on the particle size of spray-dried powders.

MATERIALS AND METHODS

1,2 distearoylphosphatidylcholine (DSPC) was received from Lipoid GmbH (Germany). Cholesterol (Ch) and egg phosphatidylcholine (PC), Dioleoyl-3trimethylammonium propane (DOTAP), trehalose dihydrate, phosphate-buffered saline (PBS) tablets were purchased from Sigma-Aldrich Company Ltd. (Poole, UK). Ethanol (99.8%, analytical grade) was from Fisher Scientific UK (Loughborough, UK).

Liposomes were generated using microfluidic mixing varying formulation and processing parameters as indicated in Table 1. The microfluidic device comprised two syringe pumps connected by PEEK tubing to a Y-junction (HPLC). Selected lipids were dissolved in the organic phase (ethanol) at and injected into one inlet of the microfluidic chip, whilst the aqueous phase (PBS; pH 7.4) is injected into the other. Hydrodynamic diameters of liposomes dispersed in PBS were assessed at 25°C by dynamic light scattering at an angle of 173° using a Zeta sizer NanoZS (Malvern Instruments Ltd, UK). **Spray**



Drying: 80mg/ml Trehalose solution was spray dried with Mini Spray Dryer B-200 (BUCHI UK Ltd) and the size, glass transition and moisture content was measured by Sympatec Helos laser diffraction, Dynamic Scattering Calorimetry and Thermogravimetric Analysis respectively.

Data analysis: 2-way Anova, Tuckey post-test

Table 1. Liposome Preparation

Formulation Para	Processing Parameters				
Lipid Composition	Organic	Aqueous	Flow rate	Total	Temperature
(molar ratio)	Solvent	Solvent	ratio	Flow	(°C)
				rate	
				(ml/min)	
DSPC:Ch:DOTAP (2:1:0.85)	Ethanol	PBS	3:1	5	20 and 55
. ,		pH7.4	(Aqueous:		
DSPC:Ch:PC(2:1:0.85)			solvent)		
. ,					
DSPC:Ch (2:1)					

RESULTS AND DISCUSSION

The three-lipid formulations showed distinct particle sizes and polydispersity at both 20°C and 55°C. At both temperatures' liposomes containing DSPC: Ch: PC had the largest size compared to DSPC: Ch DSPC: DOTAP. The liposomes produced at 20°C (Figure 1) were more heterogenous and significantly larger than the ones produced at 55°C.

These findings suggest the choice of lipid plays a vital role in determining liposome sizes which aligns with previous studies ^{7,8}. Lipid composition influence may be attributed to differences in lipid tail length, molecular shape and membrane fluidity, the packing arrangement and interactions between lipids ⁹.



Fig. 1. Liposome size distribution (solid block) polydispersity (solid line) of different formulations synthesised at 20°C. Results represent the mean \pm standard deviation of n=3 independent experiments. ns: $p>0.05^{***}p < 0.001$.

In addition, Temperature is a crucial factor that influences lipid bilayer fluidity, leading to alterations in vesicle size. Understanding this relationship is crucial to designing suitable liposomes for a specific biomedical application ¹⁰.

Table 2 shows that gas rate has the main effect on spray-dried trehalose particle size distribution (D50). A low atomization gas flow rate produced the the largest particle size regardless of other parameters.

https://doi.org/10.5920/bjpharm.1419



Fig. 2. Liposome size distribution (solid block) polydispersity (solid line) of different formulations synthesised at 55°C. Results represent the mean \pm standard deviation of n=3 independent experiments. ns: $p>0.05^{***}p < 0.001$.

Table 2. Effect of Spray Parameters on Particle size

Inlet Temperature (°C)	Feed Rate (%)	Gas Flow (mm)	D50	Tg (°C)	Percentage Yield (%)	Moisture Content (%)
160	3	25	9.27	120	43.6	2.69
160	11	55	2.10	114	72.8	2.43
160	3	55	1.60	120	79.2	1.87
110	3	55	2.09	120	81.0	3.57
110	11	25	9.09	112	87.3	3.80
135	7	40	2.67	121	77.4	3.07
110	3	25	9.77	120	56.7	2.15
160	11	25	2.50	124	56.3	2.92
110	11	55	2.82	119	77.1	5.20

CONCLUSIONS

The size of liposomes is significantly influenced by lipid composition and temperature. In the context of this study, it was observed that the particle size distribution of the spray-dried powder was primarily affected by the gas rate.

ACKNOWLEDGEMENTS

This work was supported by funding from the University of Hertfordshire

REFERENCES

- Kyriakidis, N. C., López-Cortés, A., González, E. V., Grimaldos, A. B. & Prado, E. O. SARS-CoV-2 vaccines strategies: a comprehensive review of phase 3 candidates. npj Vaccines 6, 28 (2021).
- Noh, J. Y., Jeong, H. W. & Shin, E.-C. SARS-CoV-2 mutations, vaccines, and immunity: implication of variants of concern. Signal Transduct. Target. Ther. 6, 203 (2021).
- Butler, S. E. et al. Distinct Features and Functions of Systemic and Mucosal Humoral Immunity Among SARS-CoV-2 Convalescent Individuals. Front. Immunol. 11, (2021).
- Battaglia, L. et al. Lipid nanoparticles for intranasal administration: application to nose-to-brain delivery. Expert Opin. Drug Deliv. 15, 369–378 (2018).
- Garmise, R. J., Staats, H. F. & Hickey, A. J. Novel dry powder preparations of whole inactivated influenza virus for nasal vaccination. AAPS PharmSciTech 8, 2–10 (2007).
- Luczo, J. M. et al. Intranasal powder live attenuated influenza vaccine is thermostable, immunogenic, and protective against homologous challenge in ferrets. npj Vaccines 6, 59 (2021).
- Roces, C. B. et al. Manufacturing Considerations for the Development of Lipid Nanoparticles Using Microfluidics. Pharmaceutics 12, 1095 (2020).
 Joshi, S. et al. Microfluidics based manufacture of liposomes
- simultaneously entrapping hydrophilic and lipophilic drugs. Int. J. Pharm. 514, 160–168 (2016).
- Soema, P. C., Willems, G.-J., Jiskoot, W., Amorij, J.-P. & Kersten, G. F. Predicting the influence of liposomal lipid composition on liposome size, zeta potential and liposome-induced dendritic cell maturation using a design of experiments approach. Eur. J. Pharm. Biopharm. 94, 427–435 (2015).
- Guimarães, D., Cavaco-Paulo, A. & Nogueira, E. Design of liposomes as drug delivery system for therapeutic applications. Int. J. Pharm. 601, 120571 (2021).